



The Inhibition Effects of Eugenol and Pulegone on *Stenotrophomonas maltophilia*: an Opportunistic Pathogen

Tuba BAYGAR¹, Nurdan SARAC², Aysel UGUR³, Taçnur BAYGAR⁴, Uydu BALCI²

¹Mugla Sitki Kocman University, Research Laboratories Center, Material Research Laboratory, Muğla-TURKEY

²Mugla Sitki Kocman University, Faculty of Science, Department of Biology, Muğla-TURKEY

³Gazi University, Faculty of Dentistry, Department of Basic Sciences, Section of Medical Microbiology, Ankara-TURKEY

⁴Mugla Sitki Kocman University, Faculty of Fisheries, Department of Seafood Processing Technology, Muğla-TURKEY

*Corresponding author: Dr. Tuba BAYGAR; E-mail: tubaygar@mu.edu.tr; ORCID: 0000-0002-1238-3227

How to cite: Baygar T, Sarac N, Ugur A, Baygar T, Balci U. The inhibition effects of eugenol and pulegone on *Stenotrophomonas maltophilia*: An opportunistic pathogen. Erciyes Üniv Vet Fak Derg 2019; 16(1): 29-35.

Summary: Aerobic, non-fermentative and Gram-negative *Stenotrophomonas maltophilia* is a multidrug-resistant bacilli that is known to be pathogen for human and animals. *S. maltophilia* has been isolated from different animal species and also found in a variety of environments including soil, water, and plants. *S. maltophilia*, which has the ability to form biofilms on surfaces that cause environmental problems, is resistant to many antibiotic classes such as cephalosporins, carbapenems, and aminoglycosides. Here in this study, its aimed to determine the inhibition activities of natural phenolic compounds eugenol and pulegone against *S. maltophilia* MU69. Antibacterial activities of eugenol and pulegone were initially determined by disc diffusion method. Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC) were also determined by tube dilution assay. Antibiofilm activities of the compounds were investigated by crystal violet staining and also monitored using Scanning Electron Microscopy (SEM). Zone of inhibition measurements were found to be 39 mm and 28 mm for eugenol and pulegone, respectively. MIC and MBC values were found to be 3.12 mg/mL for eugenol while these were 2.5 mg/mL and 5 mg/mL for pulegone, respectively. The highest antibiofilm activity was found to be 56.30±0.45% for MIC of eugenol. To our knowledge, this is the first report of the antibacterial and antibiofilm activity of eugenol and pulegone against *S. maltophilia*. According to the biological activity results, it can be concluded that these natural agents may be potentially used for veterinary sciences, food industry or pharmaceutical applications that aim to manage *S. maltophilia* biofilm.

Key words: Antibacterial, antibiofilm, eugenol, pulegone, *S. maltophilia*.

Eugenol ve Pulegonun Fırsatçı Patojen Bir Bakteri Olan *Stenotrophomonas maltophilia*'ya Karşı İnhibisyon Etkinliğinin Belirlenmesi

Özet: Aerobik, non-fermentatif ve gram-negatif *Stenotrophomonas maltophilia*, insan ve hayvanlar için patojen olarak bilinen çoklu ilaç direncine sahip bir basildir. Farklı hayvan türlerinden izole edilmiş *S. maltophilia* suşu toprak, su ve bitkiler gibi çeşitli çevresel etmenlerden de izole edilebilmektedir. Katı yüzeyler üzerinde biyofilm oluşturabilme yeteneği sayesinde *S. maltophilia* suşu çevre sorunlarına yol açmakla birlikte aminoglikozitler, karbapenemler ve sefalosporinler gibi çeşitli antibiyotik sınıflarına karşı direnç göstermektedir. Bu çalışmada doğal fenolik bileşikler olan eugenol ve pulegonun *S. maltophilia* MU69 suşu üzerindeki inhibisyon aktivitelerinin belirlenmesi amaçlanmıştır. Eugenol ve pulegonun antibakteriyel aktivitesi disk difüzyon yöntemi ile araştırılmıştır. Minimum İnhibisyon Konsantrasyonu (MİK) ve Minimum Bakterisidal Konsantrasyonu (MBK) ise tüp dilüsyon yöntemi ile belirlenmiştir. Antibiyofilm aktivitenin belirlenmesinde kristal viyole boyama yöntemi uygulanmış ve ayrıca Taramalı Elektron Mikroskopi (SEM) ile görüntülenmiştir. Eugenol ve pulegon için inhibisyon zonu sırasıyla 39 mm ve 28 mm olarak ölçülmüştür. MİK ve MBK değerleri eugenol için 3.12 mg/mL olarak belirlenirken, pulegon için sırasıyla 2.5 mg/mL ve 5 mg/mL olarak tespit edilmiştir. En yüksek antibiyofilm aktivite oranı % 56.30±0.45 ile eugenole ait MİK değeri için gözlenmiştir. Bu çalışma eugenol ve pulegon bileşiklerinin *S. maltophilia* suşuna karşı antibakteriyel ve antibiyofilm aktivitelerinin belirlendiği ilk çalışma olarak sunulmaktadır. Biyolojik aktivite çalışmalarının sonuçlarına göre bu iki doğal bileşik veteriner bilimlari, gıda endüstrisi ve farmasötik uygulamalarda *S. maltophilia*'ya karşı geliştirilen sistemlerde potansiyel kullanım kapasitesine sahiptir.

Anahtar kelimeler: Antibakteriyel, antibiyofilm, eugenol, pulegon, *S. maltophilia*.

Introduction

There is a growing interest in the use of herbal products as an alternative to standard medical therapies. Plants contain various type of bioactive compounds

and secondary metabolites that play an important role for using throughout alternative pharmaceutical approaches. Some of these compounds are found in essential oils isolated from plants. Essential oils have distinctive fragrances and/or flavours and are used in cosmetic as well as medical applications (38). Essential oils (EOs) are aromatic and volatile liquids ex-

tracted from plant materials, such as flowers, roots, bark, leaves, seeds, peel, fruits, wood, and whole plant (19). Essential oils (EOs) can be isolated from by several techniques such as water or steam distillation, solvent extraction, expression under pressure, supercritical fluid extraction, subcritical water extraction, ultra-sound assisted extraction, microwave assisted extraction (7). The main constituents of EOs are mono- and sesquiterpenes, along with carbohydrates, phenols, alcohols, ethers, aldehydes, and ketones, which are responsible for the biological activity of aromatic and medicinal plants as well as for their fragrance (37). Eugenol (2-allyl-4-methoxyphenol) is a phenolic compound that can be obtained from a wide range of plant sources including clove oil, nutmeg oil, cinnamon extract and many other plants (28). The pharmaceutical functions of essential oils isolated from various plants have also thought to be due to the presence of eugenol (36). Eugenol is known to be effective against a number of lifestyle related threats including nervous disorders, digestive complications, reproductive derangements, blood cholesterol irregularity, hyper-tension, elevated blood glucose level, microbial infections, inflammatory actions and carcinogenesis (45). Antioxidant (25), antimicrobial (2), anti-inflammatory (29), anticancer (43), anti-diabetic (39), neuroprotective (35), and antihypercholesterolemic (42) activities of eugenol have been reported. Pulegone [Cyclohexanone; 5-methyl-2-(1-methylethylidene)-] is a monoterpene ketone present in the leaves and flowering tops of several members of the mint family *Lamiaceae*. Pulegone is reported to be the major constituent of, *Mentha pulegium* (22), *M. longifolia* (34), and *Micromeria cilicica* (18). As a pharmacologically active compound, pulegone found to have antimicrobial activity against *Candida albicans* and *Salmonella typhimurium* (18). Gram-negative bacterial pathogens, as *Stenotrophomonas maltophilia*, are often multiple-drug-resistant organisms (MDROs) due to multidrug resistance pumps, plasmids harboring antibiotic resistance genes, and various gene transfer mechanisms involved in the acquisition of antimicrobial resistance (9). *S. maltophilia* which is generally associated with respiratory infections in humans is considered as a nosocomial bacterium (1), but there have been reports of *S. maltophilia* associated with community acquired infections (9). *S. maltophilia* was considered to be the cause of fleece rot in sheep (31), septicemia in crocodiles (23) and the snubnose dart (*Trachinotus ovatus*) (47), ulcerative stomatitis in captive snakes (17), lymphadenitis in goats (26), infectious intussusception syndrome (IIS) in channel catfish (21), gill disease in sea bream (27), and lower airway disease in horses and dogs (1). *S. maltophilia* has the ability to adhere to solid surfaces such as plastics and form bacterial films (biofilms). The drug resistance mechanisms of the *S. maltophilia* are acquired by the horizontal transfer of antibiotic re-

sistance through plasmids (5), transposons (6), integrons (24), efflux pumps, melanin-like pigment and biofilm formation (30).

The published sequence of the *S. maltophilia* genome shows numerous resistance genes, such as genes encoding for multidrug-efflux pumps, β -lactamases, and aminoglycoside-modifying enzymes (13). Discovery of the new strategies to treat the *S. maltophilia* infections has been gaining importance for a long time. In this study, its aimed to determine the antimicrobial and antibiofilm activities of eugenol and pulegone against *S. maltophilia* MU69 as a natural alternative to the use of antibiotics.

Material and Methods

Materials

Eugenol and pulegone were purchased from Sigma-Aldrich. Mueller Hinton Agar (MHA), Tryptic Soya Broth (TSB) and D-Glucose were purchased from Merck. *S. maltophilia* MU69 strain was provided from Mugla Sıtkı Kocman University Culture Collection and was incubated at $37\pm 0.1^\circ\text{C}$ for 24 h. Inocula was prepared adjusting the turbidity of the medium to match the 0.5 McFarland standard dilutions. The strain was maintained in its appropriate agar slants at 4°C throughout the study and used as stock culture. Eugenol and pulegone concentrations were prepared with 10 % DMSO as two-fold serial dilutions.

Disc Diffusion Assay of Eugenol and Pulegone

The antimicrobial activity was measured based on the disc diffusion method using pour-plating technique (12). 20 mL of MHA (Merck) sterilized and cooled to $45-50^\circ\text{C}$. After injecting 1000 μL microorganism cultures to sterile plates, media was distributed and mixed homogenously. Sterile 6 mm paper discs (Schleicher and Schuell) were impregnated with 20 μL of pure eugenol and pulegone and were then placed on the inoculated agar. The plates were incubated at appropriate temperature for performing the bacteria, as mentioned above. At the end of the incubation period, diameters of growth zones around the discs were measured. The experiments were performed in triplicate.

Tube Dilution Method

Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC) of eugenol and pulegone were determined by tube dilution method as recommended by the Clinical and Laboratory Standards Institute (11). *S. maltophilia* was grown in Mueller-Hinton Broth (MHB) at 37°C overnight and diluted to 5×10^5 colony forming unit (CFU)/mL. Eugenol/ Pulegone solutions (200 μL)+microorganism (20 μL)+culture media (1800 μL) were inoculated into a glass tubes and incubated for 24 h. 10% DMSO is

used as negative control. The MIC was defined as the lowest concentration of eugenol/pulegone showing no turbidity (no visible growth of microorganism). 100 µL cell suspensions from the tubes with no turbidity were poured onto Mueller-Hinton Agar (MHA) plates and incubated overnight and the MBC was defined as the lowest concentration of eugenol/pulegone at which bacteria did not grow on agar media. The experiments were performed in triplicate.

Effect on biofilm formation

The effect of eugenol and pulegone on biofilm-forming ability of *S. maltophilia* were tested by a microplate biofilm assay (33). Bacterial strain was prepared as above, but Trypticase Soy Broth (TSB) supplemented with 5% D-glucose was used as culture media. Cultures diluted to 1:100 in fresh TSB with 5% D-glucose. Totally 200 µL bacteria suspension and MIC and MIC/2 of eugenol and pulegone were incubated in sterile microplate at 37°C for 48 h. After incubation, the wells were washed with distilled water twice to remove the planktonic bacteria. The remaining bacteria was subsequently stained with 0.1% crystal violet solution for 10 minutes. Wells were washed again to remove the excess crystal violet solution. After air-

mentioned above. Control group was incubated without eugenol or pulegone addition. After incubation period at 37 °C for 48 h, the coverslips were gently rinsed with PBS (pH 7.4) and fixed with 2.5% glutaraldehyde at 4 °C for 2 h. Following the glutaraldehyde fixation, the coverslips were washed again with PBS for 1 h and dehydrated by increasing concentrations of ethanol. Specimens were air-dried and coated by gold (Emmitech K550, UK) before examining with a SEM (JEOL, JSM-7600F; JEOL Ltd., Tokyo, Japan).

Results

Zone of inhibition measurements were found to be 39 mm and 28 mm for eugenol and pulegone, respectively. MIC and MBC values were found to be 3.12 mg/mL (both are the same) for eugenol while these were 2.5 mg/mL and 5 mg/mL for pulegone, respectively.

The MIC value of eugenol obtained in this study was found to be same as the MBC value, which suggests that eugenol is bacteriostatic and bactericidal at 3.12 mg/mL concentration. On the other hand, pulegone was found to be bacteriostatic at lower concentration (2.5 mg/mL) than its bactericidal concentration (5 mg/

Table 1. MIC and MBC values of eugenol and pulegone against *S. maltophilia*

	Eugenol (mg/mL)	Pulegone (mg/mL)
MIC	3.12	2.50
MBC	3.12	5.00

*mg/mL: miligram/milliliters

drying, 200 µl of ethanol were added to each well and incubated at room temperature for 10 m. 125 µl solution from each well transferred to another sterile tube and the final volume was adjusted to 1 mL with distilled water. Optical density of the solutions was measured at 550 nm (Multiskan GO UV/Vis Microplate Spectrophotometer, Thermo-Fisher Scientific, USA). The effect of eugenol and pulegone on biofilm formation of *S. maltophilia* was calculated with the following equation:

$$\% \text{ Antibiofilm Effect} = \frac{[(\text{Control}_{OD} - \text{Sample}_{OD}) / \text{Control}_{OD}] * 100}$$

where control is the cell suspensions of bacteria containing 10% DMSO without eugenol/pulegone. The experiments were performed in triplicate.

Scanning Electron Microscopy (SEM)

To monitorize the antibiofilm activity, the highest biofilm inhibition was also observed by SEM using glass coverslips and compared with the control group (8). Sterile circle glass coverslips (20×20 mm) were placed in biofilm assay tubes which were prepared as

mL) (Table 1).

Within the results of this study, the highest antibiofilm activity was found to be 56.30±0.45% for MIC of eugenol (Figure 1). Inhibition effect of pulegone was slightly lower than eugenol (39.31±0.79%). Antibiofilm activity of subMIC concentrations were similar for

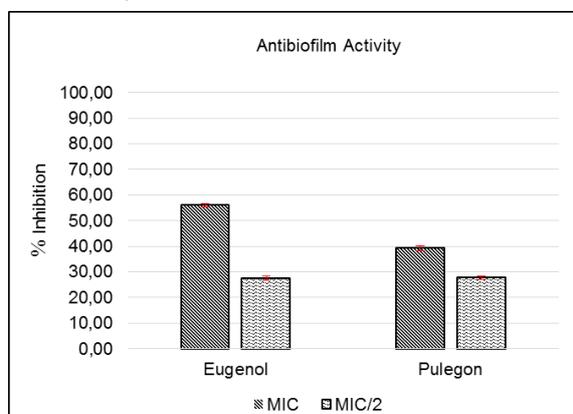


Figure 1. Antibiofilm activity of eugenol and pulegone against *S. maltophilia*. Error bars represent the standard error of the means from three biological replicates

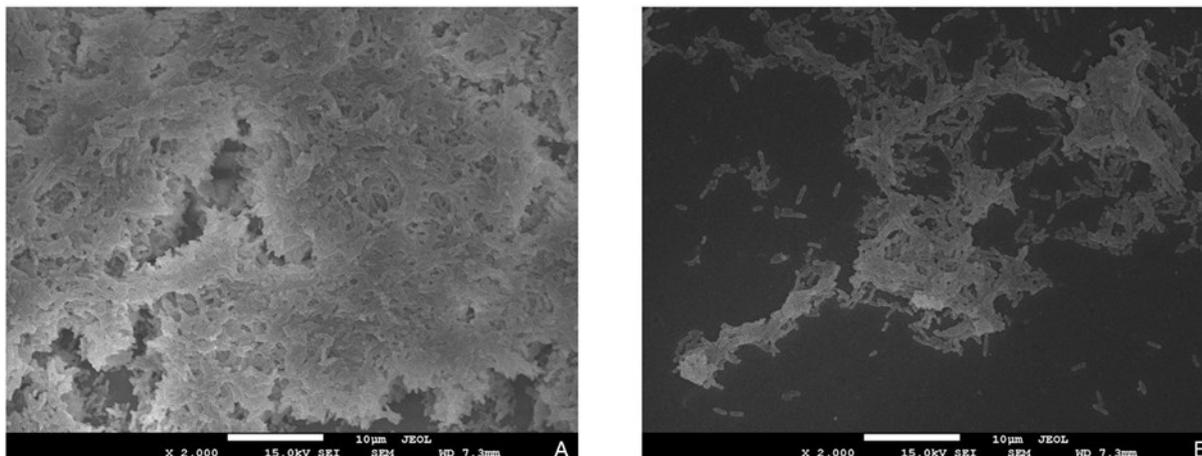


Figure 2. SEM images of *S. maltophilia* biofilm inhibition of eugenol at MIC concentration All images are taken at x2000 magnification. A: Control, B: Eugenol treatment

both eugenol and pulegone.

As the highest biofilm inhibition rate was observed for MIC of eugenol ($56.30 \pm 0.45\%$) *S. maltophilia* biofilm layer incubated with MIC of eugenol was also observed by SEM (Figure 2). The biofilm formation on the control group glass coverslips which did not contain eugenol (Figure 2A) were highly intense and it was clear that bacteria cells colonized and accumulated into the biofilm matrix. For the eugenol-incorporated group, the microbial adhesion onto the glass coverslips were found to be decreased (Figure 2B). These findings are the observable proof of the antimicrobial and antibiofilm activity of eugenol against *S. maltophilia*. As the biofilm inhibition rate was found to be lower, *S. maltophilia* biofilm formation with pulegone incorporation was not monitored using SEM.

Discussion and Conclusion

Within the results of this study, eugenol found to be more effective against *S. maltophilia* than pulegone. Similar studies revealed out that eugenol exhibits an excellent bactericidal activity against a wide range of organisms like *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa* (44), *Listeria monocytogenes* (20), and *Salmonella typhi* (16). Recent reports about the antimicrobial susceptibility of *S. maltophilia* strains revealed that trimethoprim-sulfamethoxazole (TMP-SXT), chloramphenicol, minocycline and levofloxacin are active antibiotics for the treatment of the infections caused by *S. maltophilia* (10,14). According to the literature data, there has been limited study on the antibacterial activity of eugenol and pulegone against a MDRO like *S. maltophilia*, so, the present study is an important data. MIC is the lowest concentration of the drug that allows no visible growth and MBC is defined as the lowest concentration of the drug that kills nearly 99.9% of the original inoculum (32). The MIC and MBC experi-

ments revealed the least concentrations at which eugenol and pulegone act as bacteriostatic and bactericidal agent against *S. maltophilia*, respectively. *S. maltophilia* isolates often display high-level multidrug resistance (41). Recent evidence indicates that antibiotic efflux may be a contributing factor to the intrinsic and acquired multidrug resistance of *S. maltophilia* (3). It is supposed that an effective antibiotic therapy of *S. maltophilia* infections may require the targeting of efflux mechanisms, in order to render the organism more susceptible to available antimicrobial agent (46). Bacterial biofilms are formed when planktonic organisms come together to form a community which attaches to a solid surface and encased in an exopolysaccharide matrix (32). There are a few studies about the antibiofilm activity of eugenol and pulegone for evaluating their clinical usage. Al-Shabib et al. (4) reported eugenol as a broad-spectrum anti-quorum sensing and antibiofilm agent against toxin producing biofilm forming methicillin-resistant *Staphylococcus aureus* (MRSA). In a study of Upadhyay et al. (40), it is suggested that eugenol could potentially be used to control *Listeria monocytogenes* biofilms in food processing environments. Biofilm formation by *S. maltophilia* isolates from device-associated nosocomial infections has been investigated by researchers. De Oliveira-Garcia et al. (15) suggested that flagella and fimbriae 1 (SMF-1) produced by *S. maltophilia* can be involved in biofilm formation of the bacteria. Recently, natural plant-derived extracts are investigated for their activity on eradication of the biofilm formation of microorganisms. As far as we know, this is the first study about the antibiofilm activity of eugenol and/or pulegone against *S. maltophilia* strain. According to Brooke (9), there is ongoing debate about the use of monotherapy versus combination therapy to treat infections of *S. maltophilia*, so new treatment strategies have included the use of select antibiotics in synergy. Euge-

nol or pulegone may be suggested as an alternative synergistic compounds to enhance the mechanism of antibiotics.

In conclusion, antibiotics play an important role for medical treatment. However, there is growing crisis due to the antimicrobial resistance of microorganisms against the antibiotic drugs used in the pharmaceutical applications. Researchers now focus on the investigations of naturally occurring molecules with antimicrobial and antibiofilm activity. This study results figured out that both eugenol and pulegone are highly active against *S. maltophilia* strain which has been known as multidrug-resistant bacteria. In respect to the higher inhibition zone and antibiofilm activity results, eugenol may be a clinically important compound to be tested with further analysis such as toxicity, pharmacokinetics, pharmacodynamics and drug interactions.

Acknowledgements: This study was presented at the "2nd International Congress on Advances in Veterinary Sciences and Technichs (ICAVST), October 4-8, 2017, Skopje, Macedonia" as an oral presentation.

References

- Albini S, Abril C, Franchini M, Hüsey D, Filioussis G. *Stenotrophomonas maltophilia* isolated from the airways of animals with chronic respiratory disease. *Schweiz Arch Tierheilkd* 2009; 151(7): 323-8.
- Ali SM, Khan AA, Ahmed I, Musaddiq M, Ahmed KS, Polasa H, Ahmed N. Antimicrobial activities of Eugenol and Cinnamaldehyde against the human gastric pathogen *Helicobacter pylori*. *Ann Clin Microbiol Antimicrob* 2005; 4(20): 1-7.
- Alonso A, Martinez JL. Multiple antibiotic resistance in *Stenotrophomonas maltophilia*. *Antimicrob Agents Chemother* 1997; 41:1140-2.
- Al-Shabib NA, Husain FM, Ahmad I, Baig MH. Eugenol inhibits quorum sensing and biofilm of toxigenic MRSA strains isolated from food handlers employed in Saudi Arabia. *Biotechnol Bio-technol Equip* 2017; 31(2): 387-96.
- Avison MB, Higgins CS, von Heldreich CJ, Bennett PM, Walsh TR. Plasmid location and molecular heterogeneity of the L1 and L2-lactamase genes of *Stenotrophomonas maltophilia*. *Antimicrob Agents Chemother* 2001; 45(2):413-9.
- Avison MB, von Heldreich CJ, Higgins CS, Bennett PM, Walsh TR. ATEM-2₁-lactamase encoded on an active Tn1-like transposon in the genome of a clinical isolate of *Stenotrophomonas maltophilia*. *J Antimicrob Chemother* 2000; 46(6): 879-84.
- Bassolé IHN, Juliani HR. Essential oils in combination and their antimicrobial properties. *Molecules* 2012; 17(4): 3989-4006.
- Baygar T, Ugur A. *In vitro* evaluation of antimicrobial and antibiofilm potentials of silver nanoparticles biosynthesized by *Streptomyces griseorubens*. *IET Nanobiotechnol* 2017; 11(6): 667-81.
- Brooke JS. *Stenotrophomonas maltophilia*: an emerging global opportunistic pathogen. *Clin Microbiol Rev* 2012; 25(1): 2-41.
- Chung HS, Hong SG, Kim YR, Shin KS, Whang DH, Ahn JY, Lee HS. Antimicrobial susceptibility of *Stenotrophomonas maltophilia* isolates from Korea, and the activity of antimicrobial combinations against the isolates. *J Korean Med Sci* 2013;28 (1): 62-6.
- CLSI-Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing. Sixteenth Informational Supplement. Document M100-S16, Wayne, PA: Clinical and Laboratory Standards Institute, 2011.
- Collins CH, Lyne PM, Grange JM, eds, *Microbiological Methods*, 7th ed, Butterworths, London, 1995; p.493.
- Crossman LC, Gould VC, Dow JM, Vernikos GS, Okazaki A, Sebahia M, Adlem E. The complete genome, comparative and functional analysis of *Stenotrophomonas maltophilia* reveals an organism heavily shielded by drug resistance determinants. *Genome Biol* 2008; 9(4): 1-13.
- Çikman A, Parlak M, Bayram Y, Güdücüoğlu A, Berktaş M. Antibiotics resistance of *Stenotrophomonas maltophilia* strains isolated from various clinical specimens. *Afr Health Sci* 2016; 16 (1): 149-52.
- de Oliveira-Garcia D, Dall'Agnol M, Rosales M, Azzuz A, Alcántara N, Martinez M, Girón J. Fimbriae and adherence of *Stenotrophomonas maltophilia* to epithelial cells and to abiotic surfaces. *Cell Microbiol* 2003; 5: 625-36.
- Devi KP, Nisha SA, Sakthivel R, Pandian SK. Eugenol (an essential oil of clove) acts as an antibacterial agent against *Salmonella typhi* by disrupting the cellular membrane. *J Ethnopharmacol* 2010; 130(1): 107-15.
- Draper CS, Walker RD, Lawler HE. Patterns of oral bacterial infection in captive snakes. *J Am Vet Med Assoc* 1981; 179(11): 1223-6.
- Duru ME, Öztürk M, Uğur A, Ceylan Ö. The constituents of essential oil and *in vitro* antimicrobial activity of *Micromeria cilicica* from Turkey. *J Eth-*

- nopharmacol 2004; 94(1): 43-8.
19. El Abed N, Kaabi B, Smaali MI, Chabbouh M, Habibi K, Meiri M, Ben Hadj Ahmed S. Chemical composition, antioxidant and antimicrobial activities of *Thymus capitata* essential oil with its preservative effect against *Listeria monocytogenes* inoculated in minced beef meat. *Evid Based Complement Alternat Med* 2014; 2014:1-12.
 20. Figueiras CT, Vanetti MCD. Effect of eugenol on growth and listeriolysin O production by *Listeria monocytogenes*. *Braz Arch Biol Technol* 2006; 49(3): 405-9.
 21. Geng Y, Wang K, Chen D, Huang X, He M, Yin Z. *Stenotrophomonas maltophilia*, an emerging opportunist pathogen for cultured channel catfish, *Ictalurus punctatus*, in China. *Aquaculture* 2010; 308(3): 132-5.
 22. Harborne JB, Baxter H, Moss GP. *Dictionary of Plant Toxins*. Chichester. John Wiley and Sons, 1996; p. 310-1.
 23. Harris NB, Rogers DG. Septicemia associated with *Stenotrophomonas maltophilia* in a West African dwarf crocodile (*Osteolaemus tetraspis* subsp. *tetraspis*). *J Vet Diagn Investig* 2001; 13(3): 255-8.
 24. Hu LF, Chang X, Ye Y, Wang ZX, Shao YB, Shi W, Li JB. *Stenotrophomonas maltophilia* resistance to trimethoprim/sulfamethoxazole mediated by acquisition of sul and dfrA genes in a plasmid-mediated class 1 integron. *Int J Antimicrob Agents* 2011; 37(3): 230-4.
 25. Ito M, Murakami K, Yoshino M. Antioxidant action of eugenol compounds: role of metal ion in the inhibition of lipid peroxidation. *Food Chem Toxicol* 2005; 43(3): 461-6.
 26. Johnson EH, Busaidy R, Hameed MS. An outbreak of lymphadenitis associated with *Stenotrophomonas (Xanthomonas) maltophilia* in Omani goats. *J Vet Med B* 2003; 50(2): 102-4.
 27. Kapetanovic D, Kurtovic B, Vardic I, Teskeredzic E, Teskeredzic Z. Gill disease in a gilthead sea bream (*Sparus aurata* L.). *Medycyna Wet* 2006; 62(11): 1239-41.
 28. Khalil AA, ur Rahman U, Khan MR, Sahar A, Mehmood T, Khan M. Essential oil eugenol: sources, extraction techniques and nutraceutical perspectives. *RSC Advances* 2017; 7(52): 32669-81.
 29. Kim SS, Oh OJ, Min HY, Park EJ, Kim Y, Park HJ, Lee SK (2003). Eugenol suppresses cyclooxygenase-2 expression in lipopolysaccharide-stimulated mouse macrophage RAW264. 7 cells. *Life sciences* 2003; 73(3): 337-48.
 30. Liaw SJ, Lee YL, Hsueh PR. Multidrug resistance in clinical isolates of *Stenotrophomonas maltophilia*: roles of integrons, efflux pumps, phosphoglucosyltransferase (SpgM), and melanin and biofilm formation. *Int J Antimicrob Agents* 2010; 35(2): 126-30.
 31. MacDiarmid JA, Burrell DH. Characterization of *Pseudomonas maltophilia* isolates from fleece rot. *Appl Environ Microbiol* 1986; 51(2): 346-8.
 32. Mah TFC, O'Toole GA. Mechanisms of biofilm resistance to antimicrobial agents. *Trends Microbiol* 2001; 9(1): 34-9.
 33. Merritt JH, Kadouri DE, O'Toole GA. Growing and analyzing static biofilms. *Curr Protoc Microbiol* 2005; 22(1): 1-18.
 34. Mkaddem M, Bouajila J, Ennajar M, Lebrihi A, Mathieu F, Romdhane M. Chemical composition and antimicrobial and antioxidant activities of *Mentha (longifolia* L. and *viridis*) essential oils. *J Food Sci* 2009; 74(7): 358-63.
 35. Prasad SN. Neuroprotective efficacy of eugenol and isoeugenol in acrylamide-induced neuropathy in rats: behavioral and biochemical evidence. *Neurochem Res* 2013; 38(2): 330-45.
 36. Rauscher FM, Sanders RA, Watkins JB. Effects of isoeugenol on oxidative stress pathways in normal and streptozotocin-induced diabetic rats. *J Biochem Mol Toxicol* 2001; 15(3): 159-164.
 37. Soković M, Glamočlija J, Marin PD, Brkić D, van Griensven LJ. Antibacterial effects of the essential oils of commonly consumed medicinal herbs using an *in vitro* model. *Molecules* 2010; 15(11): 7532-46.
 38. Sousa DPD, Nóbrega FF, de Lima MR, Almeida RND. Pharmacological activity of (R)-(+)-pulegone, a chemical constituent of essential oils. *Z Naturforsch C* 2011; 66(7-8): 353-9.
 39. Srinivasan S, Sathish G, Jayanthi M, Muthukumar J, Muruganathan U, Ramachandran V. Ameliorating effect of eugenol on hyperglycemia by attenuating the key enzymes of glucose metabolism in streptozotocin-induced diabetic rats. *Mol Cell Biochem* 2014; 385(1-2): 159-68.
 40. Upadhyay A, Upadhyaya I, Kollanoor-Johny A, Venkitanarayanan K. Antibiofilm effect of plant derived antimicrobials on *Listeria monocytogenes*. *Food microbiology* 2013; 36(1): 79-89.
 41. Vartivarian S, Anaissie E, Bodey G, Sprigg H, Rolston KA. Changing pattern of susceptibility of *Xanthomonas maltophilia* to antimicrobial agents:

- implications for therapy. *Antimicrob Agents Chemother* 1994; 38: 624-7.
42. Venkadeswaran K, Muralidharan AR, Annadurai T, Ruban VV, Sundararajan M, Anandhi R, Geraldine P. Antihypercholesterolemic and antioxidative potential of an extract of the plant, Piper betle, and its active constituent, eugenol, in triton WR-1339-Induced hypercholesterolemia in experimental rats. *Evid Based Complementary Altern Med* 2014; 1-11.
 43. Vidhy N, Devaraj SN. Induction of apoptosis by eugenol in human breast cancer cells. *Indian J Exp Biol* 2011; 49(11): 871-78.
 44. Walsh SE, Maillard JY, Russell AD, Catrenich CE, Charbonneau DL, Bartolo RG. Activity and mechanisms of action of selected biocidal agents on Gram-positive and-negative bacteria. *J Appl Microbiol* 2003; 94(2): 240-7.
 45. Yogalakshmi B, Viswanathan P, Anuradha CV. Investigation of antioxidant, anti-inflammatory and DNA-protective properties of eugenol in thioacetamide-induced liver injury in rats. *Toxicology* 2010; 268(3): 204-12.
 46. Zhang L, Xian-Zhi L, Poole K. Multiple Antibiotic Resistance in *Stenotrophomonas maltophilia*: Involvement of a Multidrug Efflux System. *Antimicrob Agents Chemother* 2000; 44(2): 287-93.
 47. Zhou YC, Zhu, CH, Zhang B. Isolation and prevention of the pathogen causing large scale death on *Trachinotus ovatus*. *Mar Sci* 2001; 25(4): 40-4.