

Determination of Tolerant Genotypes Against Flood Stress in Spinach

Yeşim DAL CANBAR¹⁶⁷, Musa SEYMEN², Ayşe Özgür UNCU³, Önder TÜRKMEN⁴, Banu Çiçek ARI⁵

¹Horticulture Department of Agriculture Faculty, Selcuk University, Konya, Türkiye, ² Horticulture Department of Agriculture Faculty, Selcuk University, Konya, Türkiye, ³Department of Biotechnology, Necmettin Erbakan University, Konya, Türkiye, ⁴ Horticulture Department of Agriculture Faculty, Selcuk University, Konya, Türkiye, ⁵ Horticulture Department of Agriculture Faculty, Selcuk University, Konya, Türkiye Adresi, ⁵ Horticulture Department of Agriculture Faculty, Selcuk University, Konya, Türkiye

¹ https://orcid.org/ 0000-0002-3806-6465, ² https://orcid.org/ 0000-0002-2742-137X, ³ https://orcid.org/0000-0001-6435-579X

⁴ https://orcid.org/ 0000-0003-3218-6551, ⁵ https://orcid.org/0000-0002-1578-8561

🖂: yesim.dal@selcuk.edu.tr, dalyesim@gmail.com

ABSTRACT

Abiotic stress factors generate negative effects on agricultural production daily. With the effect of global warming, the floods that have increased recently not only affected human life negatively but also caused great losses in plant development. For this reason, developing tolerant plants against flooding stress is the most critical approach reducing yield and quality losses. The present study aimed to determine the genotypes that are tolerant of flooding stress by using the agro-morphological and physiological characteristics of the commercial varieties and S5-level spinach breeding materials. In the study, 13-day flood stress was applied to 48 hybrid cultivars and 23 spinach genotypes at the S5 stage during the seedling period. As a result, in addition to the adverse effects of flood stress on plant growth, it was determined that the tolerance was different between genotypes. In the light of the results obtained, SWA0760 F1 among commercial varieties was found to be the most tolerant variety to flood stress. At the same time, genotypes 14, 9, 21, 15, 4 and 10 from breeding lines were promising genotypes that were tolerant to flooding stress. As a result, it is predicted that the inclusion of the genotypes used in the study as parents in hybrid cultivar breeding will make significant contributions to the development of tolerant cultivars against flood stress.

Ispanakta Sel Baskını Stresine Karşı Tolerant Genotiplerin Belirlenmesi

ÖZET

Abiotik stres faktörleri gün geçtikçe tarımsal yetiştiricilikteki olumsuz etkilerini artırmaktadır. Küresel ısınmanında etkisi ile son zamanlarda artan sel baskınları insan hayatını olumsuz etkilediği gibi bitki gelişiminde de büyük kayıplara neden olmaktadır. Bu sebeple son zamanlarda sel baskını stresine karşı tolerant bitkilerin geliştirilmesi verim ve kalite kayıplarını azaltmada en önemli yaklaşımdır. Mevcut çalışmada bazı ticari çeşit ve gen havuzunda bulunan bazı ıspanak ıslah materyallerinin agro-morfolojik ve fizyolojik özelliklerinden sel baskını stresine tolerant genotiplerin belirlenmesi amaçlanmıştır. Çalışmada, 48 adet hibrit çeşit ve S5 kademesinde olan 23 adet ıspanak genotiplerine fide döneminde 13 günlük sel baskını stresi uygulanmıştır. Tam sulanan kontrol bitkileri ile kıyaslanan stres koşullarında bitki gelişiminin olumsuz etkilenmesinin yanı sıra ıslah hatlarında ve ticari çeşitlerin toleranslılığının farklı olduğu tespit edilmiştir. Elde edilen sonuçlar ışığında, ticari çeşitlerden SWA0760 F1 sel baskını stresine en tolerant çeşit olarak bulunurken, ıslah hatlarından 14, 9, 21, 15, 4 ve 10 numaralı genotipler sel baskını stresine tolerant ümitvar genotipler olarak bulunmuştur. Sonuç olarak, elde edilen genotiplerin hibrit çeşit ıslahında ebeveyn olarak melezleme programlarına dahil edilmesi sel baskını stresine tolerant çeşit geliştirilmesinde önemli katkılar sağlayacağı öngörülmektedir.

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INTRODUCTION

Flooding stress has become one of the important stress factors limiting productivity in agriculture day by day (Jackson & Colmer, 2005; Hirabayashi et al., 2013). In the last 50 years, flooding stress and its destructive effects have tended to increase due to global climate change (Arnell & Liv, 2001; Hirabayashi et al., 2013; Sasidharan et al., 2018). It has been reported that this situation affects plant growth negatively, as well as severely reducing yield and quality (10-40%) (Hodgson & Chan, 1982; Bange et al., 2004; Patel et al., 2014). Flooding occurs in many parts of the world due to excessive and irregular rainfall and inadequate drainage. During flooding, oxygen and/or CO2 shortages and high levels of ethylene accumulation occur in the plant root zone (Panda et al., 2008). In the root zone, lack of oxygen causes hypoxia or anoxia around the plant tissues during flooding, causing various internal changes in plants (Ishizawa et al., 1999; Geigenberger, 2003; Bailey-Serres & Chang, 2005). Flooding can seriously impair the performance of plants due to hypoxia in the plant rhizosphere (Gibbs & Greenway, 2003; Patel et al., 2014). As a result of, metabolic activities are inhibited, and ATP production decreases (Saglio et al., 1980; Panda & Barik, 2021). Reduced ATP production restricts the energy supply for root growth; thus, plant growth slows and stops completely (Drew, 1997; Bennett, 2003). In addition, changes such as respiratory changes, leaf chlorophyll content, and photosynthetic assimilation, especially in plants whose vegetative parts are consumed, occur during the flooding stress period (Patel et al., 2014).

Spinach (Spinacia oleracea L.) is one of the most consumed vegetable types in winter, especially in the northern hemisphere. Spinach, which is an excellent climate vegetable, is grown in an open field in spring, winter, and autumn periods in sub-tropical regions. At the same time, it is cultivated in spring or autumn periods in areas where the continental climate is dominant. Since the frequency and amount of rainfall are high in these periods, it affects productivity negatively, especially in the bottomlands (Seymen, 2021). Plant growth stage, flood time and duration, flood water status, field characteristics, plant species, and differences among genotypes significantly affect flood tolerance (Kozlowski, 1997). Many plant species, including spinach, are affected by flash floods, and morpho-physiological changes result from stress. On the other hand, it has been stated that flood-tolerant plants have the capability of decreasing the negative impact rates of flooding with the multifaceted interactions of morphological, anatomical, and physiological adaptations (Kramer, 1951; Kozlowski & Pallardy, (1997). Investigating of flood tolerance mechanisms in plants will make essential contributions to productivity in agriculture (Xu et al., 2006; Singh et al., 2009). In this study, it was aimed to determine the tolerance rates of 71 spinach genotypes under flood stress conditions. This study will determine flood stress tolerance of spinach genotypes with a large gene pool for the first time. In addition to this, the tolerant status of commercially grown spinach varieties will be revealed. On the other hand, tolerant genotypes in the gene pool will be used as parents in cultivar breeding studies, allowing the development of tolerant cultivar candidates to flood stress.

MATERIALS and METHOD

The study was carried out in the glass greenhouses of Selçuk University, Faculty of Agriculture, between September 1- December 02, 2021. In the study, 48 hybrid cultivars and 23 spinach genotypes were used in level S5 (Table 1). Seeds of these genotypes were sown in plastic pots (350 cc) filled with the peat-perlite mixture. The study was planned to have two irrigation levels, control and flood, and a single harvest time. Three seeds were planted in each pot, and the seedlings were thinned by hand so that only one plant showing homogeneous growth was left when the emergence occured. Cultural processes were applied equally to all applications throughout the experiment. Irrigation was given to all pots equally at 7-day intervals by measuring with a beaker until the flooding stress was established (18 November 2021). After this date, while the normal irrigation program was continued for control applications, in stress conditions, the drainages were closed, and flooding was created by irrigation up to the potting soil. During the 13-day flooding, the water that was lost due to the daily controls was applied to the pots again. Control and stress applications of all genotypes were harvested at once on 2 December 2021. The plant parts of the harvested plants were cut, and their roots were cleaned, and they were weighed, and the fresh plant weight (g) and root fresh weight (g) were determined. The fresh weights of the plant and root parts were dried in an oven at 72 °C. Plant dry weight (g) and root dry weight (g) were determined in the samples at a constant weight. In addition, the lengths (cm) of the root parts were determined. The fresh weight of the discs from the leaf samples was taken, and their turgor

weight was determined by saturating them with water. Then, after drying the samples in an oven at 80 °C for 48 hours, their dry weights were determined, and the relative water content (%) calculations were made according to Kaya et al. (2003). Then, the discs taken from the leaves were rinsed three times with distilled water, 10 ml of water was added, and EC values were measured after shaking them in a shaker at 25 °C for 24 hours. After the same samples were kept in an autoclave at 120 °C for 20 minutes, they were cooled to 25 °C and EC measurements were made again. Then, membrane damage (%) calculations were made

according to Lutts et al. (1996). In order to determine chlorophyll a, b, and carotenoids in the leaf samples, the samples were ground in 10 ml acetone and subsequently centrifuged at 15000 rpm. Then respectively readings, were made by spectrophotometric method at 663, 652, and 470 nm, chlorophyll a and b were determined according to Lichtenthaler and Buschmann (2001), and the number of carotenoids was determined according to the Jaspars formula (Witham et al., 1971).

Table 1. The spinach genotypes and inbreeding generations in the studyTablo 1. Çalışmada kullanılan ıspanak genotipleri ve ıslah kademeleri

Genotype	Genotype code	Genotype grade	Genotype	Genotype code	Genotype grade
number			number		
1	2k-13/1-1	$\mathbf{S5}$	37	-	El Real F1
2	10b-52/3	$\mathbf{S5}$	38	-	El Salvator F1
3	15k-1/1-2	$\mathbf{S5}$	39	-	Yaman F1
4	19a-51/1-1	$\mathbf{S5}$	40	-	Hudson F1
5	19b-52/1-2	$\mathbf{S5}$	41	-	Green Gold F1
6	19c-33/1-1	$\mathbf{S5}$	42	-	Amador F1
7	19k-42/1-2	$\mathbf{S5}$	43	-	Şahmeran F1
8	31k-24/1-1	$\mathbf{S5}$	44	-	Ayaz F1
9	32b-21/1-2	$\mathbf{S5}$	45	-	Sprinter F1
10	32c-22/1-5	$\mathbf{S5}$	46	-	Green Star F1
11	48e-12/1-2	$\mathbf{S5}$	47	-	Reis F1
12	48k-42/1-5	$\mathbf{S5}$	48	-	Sardes F1
13	49c-31/2-3	$\mathbf{S5}$	49	-	Samuray F1
14	50b-51/1-1	$\mathbf{S5}$	50	-	Apollo F1
15	51a-33/1-1	$\mathbf{S5}$	51	-	Sayonora F1
16	53d-43/1-1	S5	52	-	Region F1
17	53k-12/1-3	$\mathbf{S5}$	53	-	Revere F1
18	54k-33/2-2	$\mathbf{S5}$	54	-	Spiros F1
19	55a-31/1-2	$\mathbf{S5}$	55	-	Rembrant F1
20	56k-52/2-1	S5	56	-	Shelby F1
21	58a-2/2-1	$\mathbf{S5}$	57	-	Ranchero F1
22	63k-21/2-2	S5	58	-	Matador F1
23	67c-41/1-2	S5	59	-	Nebraska F1
24	-	Tiger F1	60	-	Vena F1
25	-	Red Kitten F1	61	-	Catrina F1
26	-	Hynea F1	62	-	SV1714VC F1
27	-	Silverwhale	63	-	Matador
28	-	Racoon F1	64	-	SV1748VC F1
29	-	Parrot F1	65	-	S044 F1
30	-	Harrier F1	66	-	Midway F1
31	-	Antelope F1	67	-	SWA0760 F1
32	-	Pigeon F1	68		Java F1
33	-	Kookaburra F1	69		Aras F1
34	-	Manatee F1	70		Anemon F1
35	-	Gazelle F1	71		Anlani F1
36	-	El Tajin F1			

Percentage changes were determined by taking the numerical differences of agro-morphological and physiological parameters obtained from flooding stress and control applications of different spinach genotypes. On the other hand, principal component analyses were performed in the JMP-14 package program to reveal essential results in the interpretation of multiple values, and tolerant genotypes were tried to be determined as well as to determine the parameters that indicate significant changes between genotypes (Seymen, 2021).

RESULTS and DISCUSSION

The Effect of Flooding Stress on Agro-Morphological Parameters

It was observed that the flooding stress applied to the spinach genotypes in the present study had significant effects on agro-morphological and physiological parameters. When Table 1 is examined, the average plant fresh weight was found to be 2.844 g, while the applied flooding stress caused a 19% decrease in spinach cultivar and genotypes. While the genotype with the highest decrease in plant weight was obtained from the number 46 by 91%, the highest increase was obtained from the number 67 by 72%. It reduced the average subsoil fresh weight by 17% under flooding conditions in spinach. The genotype with the highest decrease was found in genotype 38 by 90%, while the highest increase was found in Revere F1 (53) hybrid variety by 86%. In addition, the effect of flooding stress did not cause a decrease in the plant and root fresh weights of genotypes 21, 62, 5 and 72. As in Table 2, the negative effect of flooding stress on subsoil dry weights was seen by a 58% decrease in genotype 8In addition, the genotype that experienced the least affected by flooding stress was hybrid number 61 with an increase of 75%. Likewise, flood stress caused a 22% decrease in average subsoil dry weight. Spinach is grown mainly for its fresh leaves, and the number of leaves per plant and leaf size; therefore, leaf weight determines the total yield. Flooding stress reveals the mechanism of hypoxia conditions and the formation of adventitious roots in the soil (Kawase, 1981). Due to this situation, the average plant root length was 13.27 cm, while the applied flooding stress caused a 36% decrease in the plant root length of spinach. While the much more decrease was observed in genotype 69 by 90%, the highest increase was obtained in genotype 11 by 33% (Table 2). Under flooding conditions, plant roots are in a state of hypoxia, their metabolic activities are inhibited, and ATP production is reduced (Saglio et al., 1980). Decreased ATP production restricts the energy supply for root growth, thereby reducing vegetative growth (Liao & Lin, 2001). This high variability for the above parameters can form the basis for the effective selection of superior genotypes. Rezvani et al. (2012), on the other hand, reported that the flooding stress had negative effects plant the fresh and dry weight of the saffron plant and the fresh and dry weight of the root. Likewise, Grichko and Glick (2001) reported that flooding stress had a negative effect on the fresh and dry weight of the plant part of tomatoes in their study on tomatoes.

The Effect of Flood Stress on Physiological Parameters

It has been determined that flooding stress significantly effects on relative water content (RWC) and membrane damage. When the impacts of flooding stress was evaluated in the control application, the amount of RWC decreased by 20% (average decrease) (Table 3). The genotype with the peak sensitivity was found to be 45 (Sprinter F1), with a decrease of 89%. While the amount of membrane damage did not change much in the control application during the harvest period, greater membrane damage was obtained in the flooding stress application (Table 3). Xu and Leskovar (2015) found that drought stress decreased the amount of RWC in their study similar to spinach. Seymen (2021) found that the RWC rate increased with flooding stress in his study on spinach. Yadav and Hemantaranjan (2017) demonstrated that flooding stress decreased membrane damage in mungbean. The obtained results are thought to be affected by the cumulative effect of plant species, soil, and environmental conditions from this discussion (Boyer et al., 2008; Parkash & Singh, 2020). It has also been reported that the effect of flooding stress varies between species (Seymen, 2021). On the other hand, physiological variables also cause significant changes in the type and duration of stress. Under flooding stress conditions, short-term stress conditions increase the RWC rate by causing less water uptake from the root, while in longer-term stress conditions, it causes a decrease in the amount of RWC due to membrane damage.

As a result of the present study, it was observed significant effect in the contents of chlorophyll a (chl a), chlorophyll b (chl b), and carotenoid physiological parameters obtained by flood stress. Compared to the application, significant decreases control were observed in the mean chl a, chl b and carotenoid contents under flood stress by 21%, 19% and 16%, respectively. Genotype 1 demonstrated the most significant difference in chl a and chl b contents, with a decrease of 75% and 74%, respectively. Studies have reported that the amount of photosynthetic pigments grouped as chlorophyll a, chlorophyll b and carotenoids in flooded plants vary depending on the species, the type and duration of stress, the stage of the plant's life cycle, the concentration of stress, and the difference in genotype (Foyer & Shigeoka, 2011; Zhigou & Derrick M, 2012; Tian et al., 2021). Therefore, they reported that chlorophyll content decreased due to flood stress, damage to the chloroplast membrane and its structure, photo-oxidation of chlorophyll, increased chlorophyllase activity, and suppression of chlorophyll biosynthesis (Kingston-Smith & Foyer, 2000; Kabiri et al., 2014). In this sense, the amount of chlorophyll decreased due to the deficiencies observed in leaf and plant development and flooding stress. Similarly, decreases in chlorophyll and carotene contents were observed under flood stress in spinach (Seymen, 2021), tomatoes (Ezin et al., 2010; Bhatt et al., 2015; Rasheed et al., 2018) and cabbage (Brazel et al., 2021). When the results obtained in this study were evaluated, it was seen that flood stress negatively affected the amount of chlorophyll.

Table 2.'%' changes obtained using agro-morphological traits among spinach genotypes under flood stress conditions
Tablo 2. Sel baskını stres koşulları altında ıspanak genotipleri arsında agro-morfolojik özellikler kullanılarak elde edilen '% 'değişimler

Genotype	Genotype code	Plant fresh weight(g)			Root fresh weight (g)			Plant dry weights (g)			Root	dry weights	s (g)	Root lengths (cm)		
no		Control	Flooding	Difference(%)	Control	Flooding	%	Control	Flooding	%	Control	Flooding	%	Control	Flooding	%
1	2k-13/1-1	2.90	1.88	-0.35	0.36	0.31	-0.16	0.49	0.38	-0.22	0.06	0.05	-0.05	6.33	5.83	-0.08
2	10b-52/3	2.84	2.46	-0.14	0.38	0.25	-0.33	0.42	0.36	-0.15	0.05	0.04	-0.09	7.17	5.67	-0.21
3	15k-1/1-2	3.67	2.92	-0.20	0.37	0.33	-0.10	0.58	0.47	-0.19	0.06	0.05	-0.20	7.83	5.00	-0.36
4	19a-51/1-1	3.26	3.48	0.07	0.33	0.29	-0.11	0.49	0.54	0.08	0.05	0.05	-0.06	5.58	7.25	0.30
5	19b-52/1-2	3.26	1.92	-0.41	0.24	0.24	0.00	0.49	0.32	-0.35	0.05	0.03	-0.33	6.42	4.50	-0.30
6	19c-33/1-1	2.72	1.75	-0.36	0.41	0.28	-0.31	0.46	0.35	-0.24	0.07	0.03	-0.54	7.83	3.25	-0.59
7	19k-42/1-2	2.56	1.51	-0.41	0.37	0.25	-0.33	0.28	0.28	0.00	0.05	0.03	-0.46	7.83	3.50	-0.55
8	31k-24/1-1	3.92	1.35	-0.66	0.29	0.15	-0.48	0.59	0.25	-0.58	0.06	0.03	-0.51	10.50	5.08	-0.52
9	32b-21/1-2	2.66	3.14	0.18	0.24	0.38	0.63	0.37	0.43	0.16	0.04	0.04	0.09	6.65	3.65	-0.45
10	32c-22/1-5	2.45	2.00	-0.18	0.25	0.25	0.01	0.42	0.44	0.05	0.04	0.05	0.17	8.17	5.25	-0.36
11	48e-12/1-2	3.08	2.11	-0.32	0.24	0.30	0.23	0.44	0.46	0.03	0.04	0.05	0.25	5.00	6.67	0.33
12	48k-42/1-5	2.59	2.18	-0.16	0.19	0.19	-0.05	0.38	0.45	0.17	0.03	0.03	0.01	6.88	5.75	-0.16
13	49c-31/2-3	2.45	2.96	0.21	0.18	0.31	0.77	0.37	0.46	0.25	0.03	0.06	0.82	7.25	5.00	-0.31
14	50b-51/1-1	1.60	1.57	-0.02	0.14	0.25	0.82	0.21	0.27	0.27	0.02	0.03	0.40	7.50	9.50	0.27
15	51a-33/1-1	2.92	3.23	0.11	0.39	0.44	0.12	0.47	0.57	0.21	0.06	0.08	0.37	16.00	13.08	-0.18
16	53d-43/1-1	3.20	2.66	-0.17	0.28	0.37	0.34	0.65	0.49	-0.25	0.06	0.07	0.18	5.75	7.20	0.25
17	53k-12/1-3	3.83	4.77	0.24	0.29	0.51	0.73	0.63	0.75	0.19	0.05	0.08	0.61	9.25	10.10	0.09
18	54k-33/2-2	3.84	2.73	-0.29	0.26	0.32	0.23	0.66	0.55	-0.17	0.05	0.02	-0.49	6.58	4.92	-0.25
19	55a-31/1-2	2.80	2.23	-0.20	0.26	0.23	-0.13	0.38	0.48	0.27	0.05	0.01	-0.80	6.25	3.33	-0.47
20	56k-52/2-1	2.11	1.68	-0.20	0.12	0.10	-0.17	0.32	0.28	-0.15	0.03	0.02	-0.29	5.33	4.00	-0.25
21	58a-2/2-1	2.35	2.35	0.00	0.51	0.55	0.09	0.39	0.58	0.46	0.07	0.08	0.09	19.50	7.17	-0.63
22	63k-21/2-2	3.55	2.00	-0.44	0.29	0.20	-0.31	0.53	0.52	-0.03	0.06	0.05	-0.12	9.42	5.33	-0.43
23	67c-41/1-2	4.47	2.62	-0.41	1.29	0.29	-0.77	0.65	0.54	-0.17	0.16	0.05	-0.70	25.33	5.25	-0.79
24	Tiger F1	3.81	1.18	-0.69	0.91	0.11	-0.88	0.54	0.26	-0.53	0.09	0.02	-0.82	19.80	2.83	-0.86
25	Red Kitten F1	3.40	2.36	-0.31	1.13	0.36	-0.68	0.60	0.38	-0.37	0.14	0.06	-0.57	18.20	5.58	-0.69
26	Hynea F1	2.05	1.00	-0.51	0.34	0.19	-0.43	0.36	0.23	-0.36	0.04	0.02	-0.50	17.00	21.80	0.28
27	Silverwhale	2.80	1.98	-0.29	0.86	0.35	-0.59	0.32	0.24	-0.24	0.10	0.04	-0.56	25.00	9.08	-0.64
28	Racoon F1	3.37	1.92	-0.43	1.15	0.27	-0.77	0.53	0.33	-0.38	0.12	0.04	-0.69	18.42	4.25	-0.77
29	Parrot F1	2.26	2.37	0.05	0.17	0.21	0.21	0.36	0.43	0.18	0.03	0.03	0.23	8.92	6.33	-0.29
30	Harrier F1	2.59	2.15	-0.17	0.18	0.20	0.06	0.45	0.45	0.01	0.04	0.02	-0.53	7.67	4.92	-0.36
31	Antelope F1	2.66	2.41	-0.10	0.46	0.37	-0.18	0.45	0.45	-0.01	0.09	0.06	-0.36	21.10	6.50	-0.69
32	Pigeon F1	2.32	1.99	-0.14	0.56	0.23	-0.59	0.38	0.36	-0.06	0.08	0.04	-0.50	17.92	4.67	-0.74
33	Kookaburra F1	1.81	2.33	0.29	0.41	0.18	-0.56	0.33	0.49	0.51	0.06	0.04	-0.24	15.33	4.25	-0.72
34	Manatee F1	2.63	2.16	-0.18	0.62	0.24	-0.61	0.50	0.39	-0.22	0.09	0.04	-0.56	18.92	5.75	-0.70
35	Gazelle F1	2.08	2.14	0.03	0.45	0.46	0.02	0.38	0.40	0.06	0.07	0.05	-0.18	18.42	6.92	-0.62

36	El Tajin F1	2.18	2.03	-0.07	0.75	0.37	-0.50	0.44	0.41	-0.08	0.10	0.06	-0.40	21.90	8.83	-0.60
37	El Real F1	1.79	1.31	-0.27	0.66	0.14	-0.79	0.30	0.26	-0.13	0.08	0.02	-0.72	14.08	7.08	-0.50
38	El Salvator F1	1.92	0.90	-0.53	1.41	0.14	-0.90	0.32	0.16	-0.49	0.16	0.01	-0.91	17.25	5.50	-0.68
39	Yaman F1	3.93	2.27	-0.42	0.48	0.36	-0.25	0.63	0.49	-0.23	0.09	0.06	-0.29	16.00	5.75	-0.64
40	Hudson F1	4.08	4.18	0.02	0.55	0.80	0.44	0.69	0.79	0.14	0.09	0.10	0.09	16.08	17.17	0.07
41	Green Gold F1	3.49	2.11	-0.40	0.64	0.27	-0.57	0.54	0.36	-0.33	0.08	0.04	-0.49	16.83	11.58	-0.31
42	Amador F1	2.94	2.62	-0.11	0.65	0.39	-0.40	0.50	0.49	-0.01	0.09	0.07	-0.26	18.83	8.33	-0.56
43	Şahmeran F1	3.57	0.32	-0.91	0.57	0.35	-0.39	0.67	0.56	-0.17	0.10	0.07	-0.36	12.08	10.08	-0.17
44	Ayaz F1	4.45	3.41	-0.23	0.56	0.41	-0.26	0.78	0.59	-0.24	0.10	0.09	-0.13	16.42	13.30	-0.19
45	Sprinter F1	2.64	3.10	0.17	0.30	0.51	0.69	0.48	0.51	0.06	0.06	0.06	0.03	10.92	13.25	0.21
46	Green Star F1	3.48	2.69	-0.23	0.20	0.37	0.85	0.54	0.51	-0.07	0.10	0.06	-0.40	11.08	6.58	-0.41
47	Reis F1	3.39	2.94	-0.13	0.46	0.34	-0.25	0.54	0.60	0.10	0.06	0.06	-0.08	10.30	5.60	-0.46
48	Sardes F1	1.86	1.39	-0.25	0.41	0.17	-0.58	0.32	0.25	-0.20	0.05	0.03	-0.46	14.33	4.70	-0.67
49	Samuray F1	2.31	1.22	-0.47	0.76	0.18	-0.76	0.42	0.22	-0.49	0.09	0.03	-0.68	18.20	5.58	-0.69
50	Apollo F1	2.44	2.61	0.07	0.66	0.32	-0.51	0.39	0.44	0.12	0.08	0.05	-0.43	19.10	7.00	-0.63
51	Sayonora F1	3.51	1.79	-0.49	0.58	0.21	-0.63	0.56	0.42	-0.24	0.08	0.04	-0.50	16.75	3.83	-0.77
52	Region F1	2.84	1.78	-0.37	0.53	0.31	-0.41	0.47	0.36	-0.23	0.08	0.05	-0.37	13.20	6.58	-0.50
53	Revere F1	2.86	2.71	-0.05	0.22	0.41	0.86	0.45	0.50	0.11	0.04	0.06	0.54	9.70	9.17	-0.05
54	Spiros F1	2.75	1.88	-0.32	0.19	0.27	0.39	0.43	0.40	-0.08	0.04	0.05	0.40	5.97	5.67	-0.05
55	Rembrant F1	2.76	1.98	-0.28	0.25	0.26	0.04	0.48	0.44	-0.10	0.05	0.05	-0.01	7.25	6.00	-0.17
56	Shelby F1	3.35	2.95	-0.12	0.67	0.42	-0.37	0.53	0.44	-0.18	0.08	0.06	-0.20	13.33	8.58	-0.36
57	Ranchero F1	3.25	2.89	-0.11	0.37	0.35	-0.05	0.54	0.47	-0.13	0.04	0.06	0.24	10.08	7.58	-0.25
58	Matador F1	2.73	1.89	-0.31	0.33	0.25	-0.26	0.48	0.39	-0.20	0.06	0.04	-0.30	13.80	8.17	-0.41
59	Nebraska F1	2.17	2.18	0.00	0.38	0.29	-0.23	0.39	0.40	0.05	0.06	0.05	-0.25	18.12	8.40	-0.54
60	Vena F1	2.45	3.04	0.24	0.46	0.42	-0.07	0.46	0.44	-0.05	0.06	0.08	0.32	16.42	12.83	-0.22
61	Catrina F1	1.95	3.35	0.72	0.46	0.48	0.05	0.31	0.54	0.75	0.06	0.07	0.24	12.25	15.92	0.30
62	SV1714VC F1	2.78	1.48	-0.47	0.93	0.40	-0.57	0.50	0.32	-0.36	0.11	0.06	-0.44	24.42	5.20	-0.79
63	Matador	2.17	1.46	-0.33	0.89	0.11	-0.87	0.35	0.22	-0.37	0.10	0.02	-0.81	21.75	2.25	-0.90
64	SV1748VC F1	2.68	1.86	-0.30	1.06	0.17	-0.84	0.41	0.31	-0.24	0.11	0.03	-0.75	20.25	3.92	-0.81
65	S044 F1	2.97	2.12	-0.28	1.06	0.90	-0.15	0.46	0.31	-0.33	0.13	0.09	-0.35	20.50	12.83	-0.37
66	Midway F1	3.29	2.84	-0.14	0.81	0.81	0.00	0.58	0.49	-0.15	0.11	0.10	-0.11	12.25	13.50	0.10
67	SWA0760 F1	2.78	2.80	0.01	0.56	0.65	0.16	0.46	0.41	-0.12	0.06	0.05	-0.20	12.75	9.25	-0.27
68	Java F1	2.78	2.07	-0.26	0.53	0.61	0.14	0.42	0.33	-0.21	0.07	0.08	0.10	14.25	13.00	-0.09
69	Aras F1	2.36	2.04	-0.14	0.40	0.37	-0.09	0.37	0.33	-0.10	0.07	0.05	-0.22	8.00	8.50	0.06
70	Anemon F1	2.34	1.84	-0.21	0.36	0.24	-0.33	0.37	0.31	-0.17	0.05	0.04	-0.34	15.67	7.67	-0.51
71	Anlani F1	1.86	2.26	0.21	0.26	0.25	-0.04	0.78	0.37	-0.52	0.04	0.03	-0.19	15.08	8.92	-0.41
	mean±SE	2.84±0.08	2.25 ± 0.09	-1 9± 3%	0.50±0.03	0.33±0.02	17±5%	0.47±0.01	0.41±0	- 9± 3%	0.07± 0	0.05±0	22±4%	13.33±0.66	7.43±0.43	36±4%
	SD	0.66	0.74	26%	0.29	0.16	45%	0.12	0.12	25%	0.03	0.02	37%	5.54	3.65	32%
	min.	1.60	0.32	-91%	0.12	0.10	-90%	0.21	0.16	-58%	0.02	0.01	-91%	5.00	2.25	-90%
	max.	4.47	4.77	72%	1.41	0.90	86%	0.78	0.79	75%	0.16	0.10	82%	25.33	21.80	33%

Table 3: '%' changes obtained using physiological characteristics between spinach genotypes under flood stress conditions Tablo 3: Sel baskını stres koşulları altında ıspanak genotipleri arsında fizyolojik özellikler kullanılarak elde edilen '%' değişimler

Genotype number	Genotype code	Membran damage				RWC			chl a			chl b		carotenoid			
		Control	Flooding	%	Control	Flooding	%	Control	Flooding	%	Control	Flooding	%	Control	Flooding	%	
1	2k-13/1-1	17.49	17.94	3%	104.42	43.58	-58%	22.32	5.54	-75%	14.74	3.83	-74%	3.41	1.17	-66%	
2	10b-52/3	13.62	13.77	1%	82.01	47.14	-43%	21.77	21.28	-2%	14.01	14.09	1%	3.14	3.23	3%	
3	15k-1/1-2	12.42	11.85	-5%	75.01	27.32	-64%	20.13	9.38	-53%	13.20	6.63	-50%	2.61	1.78	-32%	
4	19a-51/1-1	16.40	10.50	-36%	90.78	23.12	-75%	12.96	9.22	-29%	9.11	6.18	-32%	1.78	1.92	8%	
5	19b-52/1-2	11.24	10.09	-10%	85.76	23.18	-73%	17.31	16.42	-5%	11.45	10.96	-4%	2.24	2.73	22%	
6	19c-33/1-1	14.33	18.41	28%	88.57	66.42	-25%	26.34	8.54	-68%	18.80	5.86	-69%	5.51	1.40	-75%	
7	19k-42/1-2	11.23	17.75	58%	179.09	124.11	-31%	12.78	6.65	-48%	8.09	4.52	-44%	1.93	1.47	-24%	
8	31k-24/1-1	13.54	8.43	-38%	132.96	63.59	-52%	13.90	11.57	-17%	9.87	8.01	-19%	2.21	2.64	20%	
9	32b-21/1-2	10.99	14.58	33%	136.64	100.89	-26%	11.02	8.68	-21%	6.97	5.40	-22%	1.77	1.63	-8%	
10	32c-22/1-5	13.32	26.51	99%	133.88	63.56	-53%	8.93	9.54	7%	6.27	6.16	-2%	1.40	1.90	36%	
11	48e-12/1-2	12.19	15.71	29%	128.20	116.78	-9%	19.93	6.49	-67%	13.88	4.14	-70%	3.03	1.59	-48%	
12	48k-42/1-5	11.57	12.19	5%	117.14	152.26	30%	22.26	16.97	-24%	28.50	22.71	-20%	-0.81	-0.34	-59%	
13	49c-31/2-3	13.40	11.07	-17%	73.67	106.14	44%	23.19	18.07	-22%	31.45	22.78	-28%	-5.39	-0.33	-94%	
14	50b-51/1-1	12.48	12.99	4%	137.37	112.44	-18%	15.05	9.40	-38%	9.35	6.19	-34%	2.38	1.58	-33%	
15	51a-33/1-1	11.26	10.99	-2%	82.85	115.39	39%	15.24	16.31	7%	9.28	15.12	63%	2.20	1.24	-44%	
16	53d-43/1-1	12.17	14.89	22%	110.23	101.13	-8%	20.96	13.03	-38%	13.24	8.00	-40%	3.30	2.44	-26%	
17	53k-12/1-3	11.52	10.58	-8%	110.79	110.11	-1%	22.11	9.70	-56%	17.88	6.55	-63%	2.48	1.80	-27%	
18	54k-33/2-2	10.96	20.77	90%	124.03	85.29	-31%	19.17	16.70	-13%	12.84	10.61	-17%	2.87	3.11	8%	
19	55a-31/1-2	10.48	8.77	-16%	123.31	79.51	-36%	17.90	14.38	-20%	11.36	9.38	-17%	2.71	3.10	15%	
20	56k-52/2-1	10.73	11.03	3%	163.60	118.55	-28%	24.28	11.97	-51%	15.79	6.38	-60%	3.57	2.25	-37%	
21	58a-2/2-1	17.05	13.55	-20%	161.69	98.49	-39%	26.44	19.80	-25%	19.56	13.79	-30%	3.35	3.77	12%	
22	63k-21/2-2	15.92	15.91	0%	84.51	95.91	13%	25.20	21.06	-16%	18.64	13.90	-25%	3.13	3.22	3%	
23	67c-41/1-2	13.42	11.56	-14%	145.56	109.15	-25%	25.20	25.08	0%	17.96	19.22	7%	3.47	4.34	25%	
24	Tiger F1	11.47	11.76	3%	135.86	101.63	-25%	32.36	18.92	-42%	30.99	14.45	-53%	3.64	2.35	-35%	
25	Red Kitten F1	11.65	17.73	52%	129.15	103.00	-20%	22.50	11.56	-49%	15.23	9.26	-39%	3.29	1.88	-43%	
26	Hynea F1	10.26	12.64	23%	139.16	99.81	-28%	24.64	16.97	-31%	17.45	11.71	-33%	3.35	2.73	-18%	
27	Silverwhale	10.82	17.82	65%	132.61	90.29	-32%	26.69	15.11	-43%	20.61	12.77	-38%	3.29	2.28	-31%	
28	Racoon F1	16.08	27.75	73%	124.89	107.23	-14%	29.41	16.83	-43%	31.73	12.12	-62%	2.77	1.24	-55%	
29	Parrot F1	10.90	21.36	96%	138.96	92.65	-33%	20.69	7.57	-63%	13.90	5.30	-62%	3.25	1.49	-54%	
30	Harrier F1	10.60	20.99	98%	143.96	98.68	-31%	30.97	12.48	-60%	24.48	8.90	-64%	4.30	1.58	-63%	
31	Antelope F1	13.05	13.46	3%	129.07	122.10	-5%	12.50	10.63	-15%	8.45	7.45	-12%	2.10	1.94	-8%	
32	Pigeon F1	11.42	21.79	91%	122.58	71.01	-42%	19.89	11.49	-42%	14.18	8.03	-43%	2.91	2.52	-13%	
33	Kookaburra F1	12.60	23.82	89%	119.40	66.06	-45%	13.88	10.68	-23%	9.26	7.48	-19%	2.26	2.25	-1%	
34	Manatee F1	11.82	16.37	39%	142.24	104.25	-27%	25.45	15.06	-41%	18.21	9.94	-45%	4.06	2.85	-30%	
35	Gazelle F1	15.37	17.22	12%	120.51	81.21	-33%	21.65	11.08	-49%	14.05	6.54	-53%	3.82	2.97	-22%	

	min. max.	10,14 19,91	8.43 28.04	-52% 99%	67.68 497.38	23.12 152.26	-89% 44%	8.93 32.43	5.54 27.73	-75% 55%	5.63 32.67	3.83 22.78	-74% 74%	-5.39 5.51	-0.34 4.34	-94% 47%
	SD	2,34	4.76	<i>39%</i>	54.74	26.31	24%	5.67	4.56	<i>29%</i>	6.26	4 .19	37%	1.36	0.91	29%
	mean±SE	13±0,28	15.07±0.56	19±5%	107±6.5	80±3.12	-20±3%	19.6±0.67	14.43±0.54	21±3%	13.8±0.74	10±0.5	-19±4%	2.9±0.16	2.42±0.11	-16±3%
70 71	Anlani F1	13.23	14.48	9%	82.96	71.58	-14%	25.08	12.80	-49%	13.99	6.08	-57%	4.57	2.82	-38%
70	Anemon F1	11.13 10.78	11.08	3%	85.17	67.49	-21%	12.00 12.98	16.19	25%	6.10	8.10	33%	2.86	3.23	13%
69	Aras F1	11.09 11.19	12.17 21.63	10% 93%	76.96 75.22	70.79 56.75	-25%	12.96	10.13 19.78	-56%	6.65	10.93	-68% 64%	2.68	$\frac{2.19}{3.95}$	-33% 47%
67 68	Java F1	14.14 11.09	11.89 12.17	-16% 10%	67.68 78.98	73.60	9% -10%	16.20 22.96	12.45 10.13	-23% -56%	9.35 16.10	$6.95 \\ 5.16$	-26% -68%	$3.13 \\ 3.28$	2.47 2.19	-21% -33%
67	SWA0760 F1	12.01 14.14	12.69	-16%	81.66 67.68	80.99 73.60	-1% 9%	18.64 16.20	12.45	-23%	9.35	6.95	-26%	3.13 3.13	$2.64 \\ 2.47$	-16% -21%
66 66	S044 F1 Midway F1	10.82 12.01	$12.05 \\ 12.69$	11% 6%	80.83 81.66	72.99 80.99	-10% -1%	12.05 18.64	14.67 21.06	$\frac{22\%}{13\%}$	7.15 11.37	9.07 19.53	27% 72%	2.08 3.13	2.50 2.64	20% -16%
64 65	SV1748VC F1 S044 F1	$19.44 \\ 10.82$	12.40	-36% 11%	84.04 80.83	59.57	-29% -10%	$15.33 \\ 12.05$	$16.74 \\ 14.67$	$\frac{9\%}{22\%}$	$10.10 \\ 7.15$	$11.46 \\ 9.07$	13% 27%	$2.45 \\ 2.08$	2.97 2.50	$\frac{21\%}{20\%}$
63 64	Matador SV1748VC F1	$\begin{array}{c} 11.97\\ 19.44 \end{array}$	17.05		80.15 84.04	55.70 50.57	-31%	12.71 15.22	$\begin{array}{c} 12.48\\ 16.74 \end{array}$	-2% 9%	8.71			$2.93 \\ 2.45$	2.22 2.97	-24% 21%
62 63	SV1714VC F1 Matador	11.62	12.47	$\frac{7\%}{42\%}$	83.77	68.50 55.70	-18%	16.17	11.39	-30% -2%	10.43	$7.70 \\ 7.92$	-26% -9%	$2.49 \\ 2.93$	2.00 2.22	-20% -24%
61 69	Catrina F1	10.70	11.80	10%	90.46	73.65	-19%	17.80	20.65	16%	12.53	12.92	3%	2.69	3.18	18%
60 61	Vena F1	12.64	10.89	-14%	82.25	76.77	-7%	22.90	13.72	-40%	14.71	9.30	-37%	3.64	2.07	-43%
59 60	Nebraska F1	14.73	10.39	-29%	75.57	68.77	-9%	21.65	14.78	-32%	14.67	9.96	-32%	3.50	2.37	-32%
58	Matador F1	19.91	9.63	-52%	80.23	58.06	-28%	14.96	18.52	24%	9.47	13.97	48%	2.36	2.42	2%
57	Ranchero F1	10.14	14.27	41%	77.44	72.00	-7%	15.17	13.78	-9%	10.40	9.51	-9%	2.59	2.53	-2%
56	Shelby F1	11.62	15.22	31%	76.94	77.42	1%	21.97	22.45	2%	14.50	18.49	28%	3.58	3.22	-10%
55	Rembrant F1	11.45	12.31	7%	80.55	65.40	-19%	11.45	10.15	-11%	7.42	6.68	-10%	1.76	1.65	-6%
54	Spiros F1	10.92	12.46	14%	80.80	67.48	-16%	12.29	16.66	36%	8.36	11.20	34%	1.90	2.71	43%
53	Revere F1	13.88	10.27	-26%	76.78	69.38	-10%	19.69	7.74	-61%	12.08	5.09	-58%	2.98	1.24	-58%
52	Region F1	10.87	14.60	34%	81.77	58.24	-29%	12.18	15.64	28%	7.94	13.81	74%	1.99	1.61	-19%
51	Sayonora F1	14.76	28.04	90%	80.52	56.58	-30%	23.17	13.52	-42%	15.53	9.46	-39%	3.32	2.38	-28%
50	Apollo F1	15.71	13.75	-12%	82.24	70.32	-14%	10.83	12.18	12%	7.59	8.41	11%	1.73	1.90	10%
49	Samuray F1	13.92	23.98	72%	71.74	61.23	-15%	15.88	10.01	-37%	10.66	7.22	-32%	2.58	2.28	-12%
48	Sardes F1	14.42	26.85	86%	73.34	61.27	-16%	25.04	11.20	-55%	16.92	7.80	-54%	3.53	2.26	-36%
47	Reis F1	15.42	17.05	11%	78.12	52.23	-33%	27.36	15.03	-45%	19.93	10.03	-50%	3.98	2.75	-31%
46	Green Star F1	16.98	12.27	-28%	72.40	72.22	0%	16.38	17.39	6%	10.86	11.40	5%	2.71	2.81	4%
45	Sprinter F1	11.24	17.03	51%	497.38	55.82	-89%	20.28	13.29	-34%	13.24	9.12	-31%	2.97	2.45	-18%
40 44	Ayaz F1	15.62 15.68	14.55 15.44	-2%	77.01	64.59	-16%	24.30	12.46	-49%	16.10	8.38	-48%	4.25 3.49	1.98	-43%
42 43	Şahmeran F1	15.68 15.52	14.59	-6%	75.04 77.42	61.86	-20%	32.43	19.50 27.73	-14%	32.67	10.79 19.66	-40%	$\frac{5.02}{4.25}$	4.19	-1%
41 42	Amador F1	12.01 13.68	12.14 23.57	1% 72%	71.50 75.04	42.83 66.08	-12%	13.40 24.55	17.42 19.50	-21%	14.26	10.79	-24%	$5.10 \\ 5.02$	3.61	-27%
40 41	Green Gold F1	11.28 12.01	13.45 12.14	19%	78.15 71.50	42.83	-40%	13.48	17.81 17.42	-26% 29%	6.05	8.92	-35% 47%	$4.43 \\ 3.10$	3.72 3.81	23%
39 40	Hudson F1	10.82 11.28	$15.31 \\ 13.45$	$\frac{41\%}{19\%}$	$77.31 \\ 78.13$	$82.81 \\ 75.03$	7% -4%	$\begin{array}{c} 17.11 \\ 24.21 \end{array}$	$17.65 \\ 17.81$	3% -26%	$9.14 \\ 13.05$	$9.56 \\ 8.55$	-35%	$\begin{array}{c} 3.55 \\ 4.45 \end{array}$	$4.10 \\ 3.72$	-16%
38 39	Yaman F1	$11.14 \\ 10.82$	11.48	$\frac{3\%}{41\%}$	96.31	112.24	17%	19.76	19.23		10.35	10.81	$\frac{4\%}{5\%}$	3.97		16%
37	El Real F1 El Salvator F1	18.94	10.73	-43% 3%	109.14	133.91	23%	11.30	15.71	39% -3%	5.63	8.23	46% 4%	2.98	$3.13 \\ 3.65$	5% -8%
36	El Tajin F1	12.31	12.08	-2%	135.35	122.41	-10%	22.80	18.68	-18%	14.59	11.21	-23%	3.79	3.28	-14%

PcA Analysis

PCA has been reported to be a very practical statistical analysis as it simultaneously analyzes quantitative and qualitative features and is used by many researchers (Dehghani et al., 2008; Eftekhari et al., 2010; Everitt & Dunn, 2010; Sabaghnia et al., 2011). In the current study, when we examined the genotypes one by one in terms of agro-morphological and physiological parameters, the study was explained in two components. According to the evaluated criteria, PC produces the critical value of each factor called "Eigenvalue", "Eigenvalue" values greater than one were taken into (Brejda et al., 2000) account and helped to reduce data complexity in two factors. In this study, the variations among spinach genotypes are explained in two components, with PC1= %34.31 and PC2= %25.59, and the principal component (PC) representing approximately 59.90% of the total variability. In order to use PC analysis in a study, it has been reported that more than 25% of the variance of the first two components should be enough to explain variations among factors (Mohammadi & Prasanna, 2003; Mozafari et al., 2019; Seymen et al., 2019). Generally, it has been stated that the first component contributes to the maximum variance, while the rest of the factors justify the remaining amount of variance (Fereidoonfar et al., 2018). Therefore, the PC analysis was found to strongly explain the study.

Using the PC1 and PC2 components, a Biplot plot was created to examine the relationship between the morphological and physiological observations-based parameters of the genotypes affected by flood stress. The degree of association between the two traits is based on the multiple traits being compared between genotypes and the identification of genotypes that can be used as parents in breeding programs because they are particularly good in certain respects (Figure 1). According to this biplot, the cosine of the angles formed by the vectors connected to each variable and represents the degree of relationship among variable (Yan & Rajcan, 2002; Dehghani et al., 2008). In this context, if the angle between the vectors in the figure is $<90^{\circ}$, there is a positive relationship, and if it is >90^o, there is a negative relationship, if the angle between the vectors is 180°, it has been reported that there is no significant relationship (Yan & Kang, 2002). Angles formed by biplot analysis under flooding stress conditions showed a high correlation between UDW, UFW, AFW, ADW and RL. Therefore, the higher AFW, the higher ADW was found. A strong correlation was found between plant dry weight after flood stress and plant survival under critical flooding conditions. Flooding inhibits new leaf formation, reducing the total leaf area and promoting leaf senescence (Kato et al., 2014). In the light of this information, genotypes 67 and 14 were found to be highly tolerant in terms of growth parameters under flooding stress conditions. Singh et al. (2014) observed that the dry weight of leaves decreased sharply by 70% in varieties sensitive to flood stress, while it decreased by only 30-40% in tolerant varieties under 17-day flood stress. A similar study was conducted by Arif et al. (2013) and Sabaghnia et al. (2015) and show the relationship between different morphological features. Moreover, it was determined that the higher the chl a and chl b values, the higher the carotene content. Therefore, a highly positive relationship was found between chl a, chl b, and carotene. Singh et al. (2014) also stated that maintaining integrity of chlorophyll content during and after flooding is essential for plant survival as it aids photosynthesis continuity under flooding and aids faster recovery in case of flooding. Accordingly, it can be stated that genotypes 2, 53, 59, and 75 are more tolerant to flooding stress in terms of chlorophyll content (Figure 1). These results support studies evaluating the correlation between agricultural variables in similar studies (Sarkar et al., 2006; Seymen, 2021). A significant negative correlation was found between RWC and membrane damage among spinach genotypes. Membrane damage content is significantly higher in susceptible variety during flooding and after exposure to air (Kawano et al., 2002; Damanik et al., 2012; Panda & Sarkar, 2013). For this reason, the genotypes that revealed the most membrane damage are less productive. In this context, genotypes 58 and 60 were determined to be highly sensitive to flooding stress.

The biplot plot can provide knowledge to the plant breeder with flexibility in finding the number of plants to be evaluated (Yan & Rajcan, 2002). The plant breeder can use multivariate methods by first identifying the combination of traits that make up an ideal genotype. In this context, it can be reported that genotype 67 (SWA0760 F1) is the most ideal line to be used in spinach breeding studies. Besides, genotypes 14, 9, 21, 15, 4, and 10 belong to the S5 level and were found to be flooding stress tolerant. In the study, it was determined that the genotypes 24, 25, 26, 27, 28, 52, 54 and 69 were the most sensitive to flooding stress (Figure 1). It can be stated that the determined genotypes can be ignored in future studies. This research was conducted for only one growing season and thus genotype-environment interactions may cause some fluctuates, but the results showed that multidimensional methods can be sufficiently informative in the selection of breeding directions.

CONCLUSION

It was determined that the tolerance levels of 23 breeding materials and 48 commercial spinach varieties under different flood stress conditions. In addition to the negative effects of the 13-day flood stress applied on the spinach seedling period, it was observed from the agro-morphological and physiological parameters that the tolerance of the breeding lines and commercial varieties were different. As a result of PCA, commercial cultivar SWA0760 F1 (genotype 67) was found to be the most tolerant line against flooding stress. Other cultivars were found to be sensitive or moderately tolerant. In this context, the development of tolerant varieties against flood stress in spinach is among the important issues. The spinach genotypes 14, 9, 21, 15, 4 and 10 in the S5 level were determined to be tolerant lines against flood stress. It is thought that these genotypes will be used as parents in hybrid breeding and included in hybridization programs and will give important results in the development of flood stress-tolerant variety. The obtained variety and variety candidates will contribute to the reduction of yield and quality losses in the agricultural lands where spinach is grown, due to the flooding stress, which increases its negative effects day by day.





Şekil 1. Sel baskını stres koşulları altında ıspanak genotipleri arsında agro-morfolojik özellikler kullanılarak PCA'dan elde edilen PC 1 ve 2'ye dayalı biplot grafiği

Researchers' Contribution Rate Statement Summary

The authors declare that they have contributed equally to the article.

Conflict of Interest Statement

The article authors declare that they do not have any conflict of interest.

REFERENCES

Arif, M., Jatoi, S. A., Rafique, T. & Ghafoor, A. (2013). Genetic divergence in indigenous spinach genetic resources for agronomic performance and implication of multivariate analyses for future selection criteria. J Sci Technol Dev, 32(1), 7-15.

Arnell, N.W. & Liv, C. (2001). Hydrology and water

resources. In: McCarthy, J.J., Canziani, O.F., Leary, N.A., Dokken, D.J., White, K.S. (Eds.), IPPC Climate Change 2001: Impacts, Adaptation and Vulnerability. Cambridge University Press, Cambridge, pp. 191–233

- Bailey-Serres, J. & Chang, R. (2005). Sensing and signalling in response to oxygen deprivation in plants and other organisms. *Annals of botany*, 96(4), 507-518.
- Bange, M., Milroy, S. & Thongbai, P. (2004). Growth and yield of cotton in response to waterlogging. *Field Crops Research*, 88(2-3), 129-142.
- Bennett, J. (2003). Opportunities for increasing water productivity of CGIAR crops through plant breeding and molecular biology. Water productivity in agriculture: limits and opportunities for improvement (pp. 103-126). Wallingford UK: CABI publishing.
- Bhatt, R. M., Upreti, K. K., Divya, M., Bhat, S., Pavithra, C. & Sadashiva, A. (2015). Interspecific grafting to enhance physiological resilience to flooding stress in tomato (*Solanum lycopersicum* L.). *Scientia Horticulturae*, 182, 8-17.
- Boyer, J. S., James, R. A., Munns, R., Condon, T. A. & Passioura, J. B. (2008). Osmotic adjustment leads to anomalously low estimates of relative water content in wheat and barley. *Functional Plant Biology*, 35(11), 1172-1182.
- Brazel, S., Barickman, T. & Sams, C. (2021). Shortterm waterlogging of kale (*Brassica oleracea* L. var. *acephala*) plants causes a decrease in carotenoids and chlorophylls while increasing nutritionally important glucosinolates. In VIII International Symposium on Human Health Effects of Fruits and Vegetables-FAVHEALTH 2021 1329 (pp. 175-180).
- Brejda, J. J., Moorman, T. B., Karlen, D. L. & Dao, T. H. (2000). Identification of regional soil quality factors and indicators I. Central and Southern High Plains. Soil Science Society of America Journal, 64(6), 2115-2124.
- Damanik, R. I., Ismail, M. R., Shamsuddin, Z., Othman, S., Zain, A. M. & Maziah, M. (2012).
 Response of antioxidant systems in oxygen deprived suspension cultures of rice (*Oryza sativa* L.). *Plant Growth Regulation*, 67(1), 83-92.
- Dehghani, H., Omidi, H. & Sabaghnia, N. (2008). Graphic analysis of trait relations of rapeseed using the biplot method. *Agronomy Journal*, *100*(5), 1443-1449.
- Drew, M. C. (1997). Oxygen deficiency and root metabolism: injury and acclimation under hypoxia and anoxia. *Annual review of plant biology*, 48(1), 223-250.
- Eftekhari, S. A., Hasandokht, M. R., Moghadam, M. R. F. F. & Kashi, A. (2010). Genetic diversity of some Iranian spinach (*Spinacia oleracea* L.) landraces using morphological traits. *Iranian Journal of Horticultural Science*, 41(1), 83-93.

- Everitt, B. S. & Dunn, G. (2010). Applied Multivariate Data Analysis, 2nd Edition. 354.
- Ezin, V., Pena, R. D. L. & Ahanchede, A. (2010). Flooding tolerance of tomato genotypes during vegetative and reproductive stages. *Brazilian Journal of Plant Physiology*, 22, 131-142.
- Fereidoonfar, H., Salehi-Arjmand, H., Khadivi, A. & Akramian, M. (2018). Morphological variability of sumac (*Rhus coriaria* L.) germplasm using multivariate analysis. *Industrial Crops and Products*, 120, 162-170.
- Foyer, C. H. & Shigeoka, S. (2011). Understanding oxidative stress and antioxidant functions to enhance photosynthesis. *Plant Physiology*, 155(1), 93-100.
- Geigenberger, P. (2003). Response of plant metabolism to too little oxygen. Current opinion in plant biology, $\theta(3)$, 247-256.
- Gibbs, J. & Greenway, H. (2003). Mechanisms of anoxia tolerance in plants. I. Growth, survival and anaerobic catabolism. *Functional Plant Biology*, 30(1), 1-47.
- Grichko, V. P. & Glick, B. R. (2001). Amelioration of flooding stress by ACC deaminase-containingplant growth-promoting bacteria. *Plant Physiology and Biochemistry*, 39(1), 11-17.
- Hirabayashi, Y., Mahendran, R., Koirala, S., Konoshima, L., Yamazaki, D., Watanabe, S., Kim, H. & Kanae, S. (2013). Global flood risk under climate change. *Nature climate change*, 3(9), 816-821.
- Hodgson, A. & Chan, K. (1982). The effect of short-term waterlogging during furrow irrigation of cotton in a cracking grey clay. *Australian Journal of Agricultural Research*, 33(1), 109-116.
- Ishizawa, K., Murakami, S., Kawakami, Y. & Kuramochi, H. (1999). Growth and energy status of arrowhead tubers, pondweed turions and rice seedlings under anoxic conditions. *Plant, Cell & Environment, 22*(5), 505-514.
- Jackson, M. & Colmer, T. (2005). Response and adaptation by plants to flooding stress. Annals of botany, 96(4), 501-505.
- Kabiri, R., Nasibi, F. & Farahbakhsh, H. (2014). Effect of exogenous salicylic acid on some physiological parameters and alleviation of drought stress in Nigella sativa plant under hydroponic culture. *Plant Protection Science*, 50(1), 43-51.
- Kato, Y., Collard, B. C. Y., Septiningsih, E. M. & Ismail, A. M. (2014). Physiological analyses of traits associated with tolerance of long-term partial submergence in rice. *AoB Plants, 6.* https://doi.org/10.1093/aobpla/plu058.
- Kawano, N., Ella, E., Ito, O., Yamauchi, Y. & Tanaka, K. (2002). Metabolic changes in rice seedlings with different submergence tolerance after desubmergence. *Environmental and Experimental Botany*, 47(3), 195-203.

- Kawase, M. (1981). Anatomical and morphological adaptation of plants to waterlogging. *Hort. Sci.* 16, 8-12.
- Kaya, C., Higgs, D., Ince, F., Amador, B. M., Cakir, A. & Sakar, E. (2003). Ameliorative effects of potassium phosphate on salt-stressed pepper and cucumber. *Journal of plant nutrition*, 26(4), 807-820.
- Kingston-Smith, A. & Foyer, C. (2000). Bundle sheath proteins are more sensitive to oxidative damage than those of the mesophyll in maize leaves exposed to paraquat or low temperatures. *Journal of experimental botany*, *51*(342), 123-130.
- Kozlowski, T. (1997). Responses of woody plants to flooding and salinity. *Tree physiology*, 17(7), 490-490.
- Kozlowski, T. T. & Pallardy, S. G. (1997). Growth control in woody plants. Elsevier.
- Kramer, P. J. (1951). Causes of injury to plants resulting from flooding of the soil. *Plant Physiology*, 26(4), 722.
- Liao, C.-T. & Lin, C.-H. (2001). Physiological adaptation of crop plants to flooding stress. *Proceedings of the National Science Council, Republic of China. Part B, Life Sciences, 25*(3), 148-157.
- Lichtenthaler, H. K. & Buschmann, C. (2001). Extraction of phtosynthetic tissues: chlorophylls and carotenoids. *Current protocols in food analytical chemistry*, 1(1), F4. 2.1-F4. 2.6.
- Lutts, S., Kinet, J. M. & Bouharmont, J. (1996). NaClinduced senescence in leaves of rice (*Oryza sativa* L.) cultivars differing in salinity resistance. *Annals* of botany, 78(3), 389-398.
- Mohammadi, S. A. & Prasanna, B. (2003). Analysis of genetic diversity in crop plants—salient statistical tools and considerations. *Crop Science*, 43(4), 1235-1248.
- Mozafari, A.-a., Ghaderi, N., Havas, F. & Dedejani, S. (2019). Comparative investigation of structural relationships among morpho-physiological and biochemical properties of strawberry (Fragaria× ananassa Duch.) under drought and salinity stresses: A study based on in vitro culture. *Scientia Horticulturae*, 256, 108601.
- Panda, D. & Barik, J. (2021). Flooding tolerance in rice: Focus on mechanisms and approaches. *Rice Science*, 28(1), 43-57.
- Panda, D. & Sarkar, R. K. (2013). Characterization of leaf gas exchange and anti-oxidant defense of rice (*Oryza sativa* L.) cultivars differing in submergence tolerance owing to complete submergence and consequent re-aeration. *Agricultural Research*, 2(4), 301-308.
- Panda, D., Sharma, S. G. & Sarkar, R. K. (2008). Chlorophyll fluorescence parameters, CO2 photosynthetic rate and regeneration capacity as a result of complete submergence and subsequent re-

emergence in rice (Oryza sativa L.). Aquatic Botany, 88(2), 127-133.

- Parkash, V. & Singh, S. (2020). A review on potential plant-based water stress indicators for vegetable crops. *Sustainability*, 12(10), 3945.
- Patel, P. K., Singh, A. K., Tripathi, N., Yadav, D. & Hemantaranjan, A. (2014). Flooding: abiotic constraint limiting vegetable productivity. *Advances in Plants and Agriculture Research*, 1(3), 96-103.https://doi.org/10.15406/apar.2014.01.00016
- Rasheed, R., Iqbal, M., Ashraf, M. A., Hussain, I., Shafiq, F., Yousaf, A. & Zaheer, A. (2018). Glycine betaine counteracts the inhibitory effects of waterlogging on growth, photosynthetic pigments, oxidative defence system, nutrient composition, and fruit quality in tomato. *The Journal of Horticultural Science and Biotechnology*, 93(4), 385-391.
- Rezvani, N., Sorooshzadeh, A. & Farhadi, N. (2012). Effect of nano-silver on growth of saffron in flooding stress. World Acad Sci Eng Technol, 1, 517-522.
- Sabaghnia, N., Asadi-Gharneh, H. & Janmohammadi, M. (2015). Genetic diversity of spinach (*Spinacia* oleracea L.) landraces collected in Iran using some morphological traits. Acta agriculturae Slovenica, 103(1), 101-111.
- Sabaghnia, N., Dehghani, H., Alizadeh, B. & Moghaddam, M. (2011). Yield analysis of rapeseed (Brassica napus L.) under water-stress conditions using GGE biplot methodology. *Journal of Crop Improvement*, 25(1), 26-45.
- Saglio, P. H., Raymond, P. & Pradet, A. (1980). Metabolic activity and energy charge of excised maize root tips under anoxia: control by soluble sugars. *Plant Physiology*, 66(6), 1053-1057.
- Sarkar, R. K., Reddy, J. N., Sharma, S. G. & Ismail, A. M. (2006). Physiological basis of submergence tolerance in rice and implications for crop improvement. *Current Science*, 91(7), 899–906. http://www.jstor.org/stable/24094287
- Sasidharan, R., Hartman, S., Liu, Z., Martopawiro, S., Sajeev, N., van Veen, H., Yeung, E. & Voesenek, L. A. (2018). Signal dynamics and interactions during flooding stress. *Plant Physiology*, 176(2), 1106-1117.
- Seymen, M. (2021). How does the flooding stress occurring in different harvest times affect the morpho-physiological and biochemical characteristics of spinach? *Scientia Horticulturae*, 275, 109713.
- Seymen, M., Yavuz, D., Dursun, A., Kurtar, E. S. & Türkmen, Ö. (2019). Identification of droughttolerant pumpkin (*Cucurbita pepo* L.) genotypes associated with certain fruit characteristics, seed yield, and quality. *Agricultural Water Management*, 221, 150-159.
- Singh, S., Mackill, D. J. & Ismail, A. M. (2009). Responses of SUB1 rice introgression lines to submergence in the field: yield and grain quality.

Field Crops Research, 113(1), 12-23.

- Singh, S., Mackill, D. J., & Ismail, A. M. (2014). Physiological basis of tolerance to complete submergence in rice involves genetic factors in addition to the SUB1 gene. *AoB Plants, 6.* https://doi.org/10.1093/aobpla/plu060.
- Tian, G., Qi, D., Zhu, J. & Xu, Y. (2021). Effects of nitrogen fertilizer rates and waterlogging on leaf physiological characteristics and grain yield of maize. Archives of Agronomy and Soil Science, 67(7), 863-875.
- Witham, F. H., Blaydes, D. F. & Devlin, R.M. (1971). Experiments in plant physiology. Van Nostrand Reinhold Compan, New York, USA, pp 55–56.
- Xu, C., & Leskovar, D. I. (2015). Effects of A. nodosum seaweed extracts on spinach growth, physiology and nutrition value under drought stress. *Scientia Horticulturae*, 183, 39-47.
- Xu, K., Xu, X., Fukao, T., Canlas, P., Maghirang-Rodriguez, R., Heuer, S., Ismail, A. M., Bailey-Serres, J., Ronald, P. C. & Mackill, D. J. (2006). Sub1A is an ethylene-response-factor-like gene that

confers submergence tolerance to rice. *Nature*, 442(7103), 705-708.

- Yadav, D.K. & Hemantaranjan, A. (2017). Mitigating effects of paclobutrazol on flooding stress damage by shifting biochemical and antioxidant defense mechanisms in mungbean (Vigna radiata L.) at preflowering stage. Legume Research: An International Journal, 40(3), 453-461. https:// doi.org/10.18805/lr.v0i0.7593
- Yan, W. & Kang, M. S. (2002). GGE biplot analysis: A graphical tool for breeders, geneticists, and agronomists. *CRC press*.
- Yan, W. & Rajcan, I. (2002). Biplot analysis of test sites and trait relations of soybean in Ontario. Crop Science, 42(1), 11-20.
- Zhigou, Z. & Derrick M, O. (2012). Physiological Mechanism of Nitrogen Mediating Cotton (Gossypium hirsutum L.) Seedlings Growth under Water-Stress Conditions. American Journal of Plant Sciences, 3(6), 721-730. http://dx.doi.org/ 10.4236/ajps.2012.36087.